by Tao Shen^a), Guo-Hui Li^b), Qing-Qing Zhong^c), Shu-Qi Wang^a), Dong-Mei Ren^a), Hong-Xiang Lou^{*a}), and Xiao-Ning Wang^{*a})

^a) Key Lab of Chemical Biology (MOE), School of Pharmaceutical Sciences, Shandong University, Jinan, P. R. China (phone: +86-531-88382012; fax: +86-531-88382548; e-mail: wangxn@sdu.edu.cn)
 ^b) Department of Pharmacy, Jinan Maternity and Child Care Hospital, Jinan, P. R. China
 ^c) School of Pharmaceutical Sciences, Shandong University of Traditional Chinese Medicine, Jinan, P. R. China

Two new germacrane-type sesquiterpenoids bearing an epoxy ring, myrrhanolide D (1) and myrrhasin A (2), together with eight known compounds, 3-10, were isolated from the resinous exudates of *Commiphora opobalsamum*. Their structures were elucidated based on the analyses of their spectroscopic data. The isolated compounds 1, 2, 6, and 8 were evaluated for their cytotoxic activities against human prostate cancer DU145 and PC3 cells.

Introduction. – The genus *Commiphora* of the Burseraceae family comprises over 150 species, and mainly occurs in Eastern Africa, Arabia, and India [1]. The resinous exudates of plants from this genus are used in indigenous medicines for the treatment of wounds, pain, arthritis, fractures, obesity, parasitic infection, and gastrointestinal diseases. Many metabolites, including terpenoids, steroids, flavonoids, lignans, and carbohydrates, have been discovered in this genus [2][3]. Isolated ingredients and crude extracts exhibited diverse biological features, such as anti-inflammatory, analgesic, cytotoxic, antimicrobial, cardiac protection, and hepatoprotective activities [1][2][4][5].

In our systematic investigation of chemical constituents from the genus *Commiphora*, *ca.* 50 secondary metabolites, including 23 new compounds, have been reported from three *Commiphora* species [6-10]. Motivated by the recent discovery of new natural products from this genus [11-14], we reinvestigated the resinous exudates of *C. opobalsamum*, and isolated two new germacrane-type sesquiterpenoids, **1** and **2**, together with eight known compounds **3**–**10**. Compounds **1**, **2**, **6**, and **8** were evaluated for their cytotoxic activities against human prostate cancer DU145 and PC3 cells.

Results and Discussion. – Myrrhanolide D (1), obtained as yellow oil, exhibited a *pseudo*-molecular-ion peak at m/z 373.2 ($[M + Na]^+$) in the positive-ion mode ESI-MS, consistent with the molecular formula $C_{18}H_{22}O_7$ deduced from the HR-ESI-MS m/z 351.1454 ($[M + H]^+$; calc. 351.1444). The ¹³C-NMR displayed 18 signals (*Table*), which were classified to correspond to five Me groups, one sp³ CH₂ group, four sp³ CH groups (three oxygenated (δ (C) 76.3, 78.5, and 88.6)), one quaternary C-atom, two C=C bonds, and three C=O groups (δ (C) 169.3, 170.2, and 199.6), based on the chemical

^{© 2014} Verlag Helvetica Chimica Acta AG, Zürich



shifts and HMQC correlations. The ¹H-NMR spectrum (Table) revealed the presence of four tertiary Me groups (δ (H) 1.86, 1.87, 2.08, and 3.41), a secondary Me group $(\delta(H) 1.16 (d, J = 6.0))$, one olefinic H-atom $(\delta(H) 5.41)$, and three CH–O groups Hatoms geminal to O-atoms ($\delta(H)$ 3.76, 5.21, and 5.10). These NMR data suggested that 1 might be a germacrane-type sesquiterpenoid with a MeO group (δ (H) 3.41; δ (C) 57.4) and an AcO group ($\delta(H)$ 2.08; $\delta(C)$ 20.1, 170.2), which was established by the analysis of its 2D-NMR spectra. The ¹H,¹H-COSY spectrum led to the establishment of structural fragments as depicted with bold lines in Fig. 1. Interpretation of long-range HMBCs Me(13)/C(7), C(11), and C(12); Me(14)/C(1), C(9), and C(10); H-C(9)/ C(8); Me(15)/C(3), C(4), and C(5); and H-C(5)/C(6) (Fig. 1) indicated that 1 possessed the same skeleton as the known sesquiterpenoid 1β , 8β -epoxy- 2α -methoxy-6- ∞ oxogermacra-9,7(11)-dien-8,12-olide [11]. The positions of the MeO group at C(2) and of the AcO group at C(5) were deduced from the cross-peaks H–C(2)/ δ (C) 57.4 (MeO) and H–C(5)/ δ (C) 170.2. The relative configuration of **1** was established by a NOESY experiment (Fig. 1), in which the correlations H-C(1)/H-C(2) and Me(14); H-C(2)/ H_{β} -C(3), H-C(5), and Me(15); H-C(5)/Me(15) indicated that H-C(1), H-C(5), Me(15), and the MeO group at C(2) were β -, β -, β -, and α -oriented, respectively. Therefore, the structure of 1 was established as 5α -acetoxy- 2α -methoxy-6-oxo-1.8epoxygermacra-7(11),9-dien-6-oxo-8,12-olide and named myrrhanolide D.

Myrrhasin A (2) was isolated as yellow oil, which displayed a pseudo molecular ion peak at m/z 329.5 ($[M + Na]^+$) in the positive-ion mode ESI-MS. Its molecular formula was determined as $C_{17}H_{22}O_5$ from the $[M + H]^+$ ion peak at m/z 307.1552 (calc.

Position	1		2	
	$\delta(H)$	$\delta(C)$	$\overline{\delta(\mathrm{H})}$	$\delta(C)$
1	5.21 (br. s)	88.6	3.27 (d, J = 11.4)	67.9
2	3.76 (br. s)	76.3	$1.87 - 1.92 (m, H_a),$	26.3
			$1.43 - 1.47 \ (m, H_{\beta})$	
3	1.62 (br. s)	31.4	$1.50 - 1.55 (m, H_a),$	29.0
	1.62 (br. s)		$1.75 - 1.80 \ (m, H_{\beta})$	
4	2.40 (br. s)	32.6	2.40-2.45(m)	33.0
5	5.10 (d, J = 9.0)	78.5	5.88 (br. s)	78.2
6		199.6		192.0
7		152.3		121.2
8		115.5		156.4
9	5.41(s)	121.7	$3.61 (d, J = 15.0, H_a), 2.98 (d, J = 15.0, H_b)$	38.3
10		146.5		58.9
11		134.1		123.6
12		169.3	7.11 (s)	138.7
Me(13)	1.87(s)	9.9	2.15(s)	10.4
Me(14)	1.86(s)	13.1	1.11 (s)	18.0
Me(15)	1.16 (d, J = 6.0)	16.1	1.02 (d, J = 6.6)	15.4
AcO	2.08 (s)	20.1, 170.2	2.19 (s)	20.6, 171.2
MeO	3.41 (s)	57.4		

Table. ¹*H*- and ¹³*C*-*NMR Data* (600 and 150 MHz, resp., in CDCl₃) of Compounds **1** and **2**. δ in ppm, *J* in Hz. Atom numbering as indicated in the Formulae.



Fig. 1. Key ${}^{1}H,{}^{1}H$ -COSY (—), HMBC (H \rightarrow C), and NOESY (H \leftrightarrow H) correlations of 1

307.1545) in the HR-ESI-MS. In the ¹³C-NMR spectrum (*Table*), we observed 17 signals. They were attributed to four Me, three sp³ CH₂, and three sp³ CH groups, one quaternary C-atom, two C=C bonds, and two C=O groups, with the aid of HMQC spectrum. The signals at δ (C) 121.2, 156.4, 123.6, and 138.7 were characteristic of furan ring C-atoms (C(7), C(8), C(11), and C(12)) in the skeleton of furanosesquterpenoids [15][16]. Consistent with the ¹³C-NMR data, the ¹H-NMR spectrum revealed the presence of four Me groups and one olefinic H-atom (δ (H) 7.11) in the furan ring. Furthermore, the signals at δ (C) 20.6 and 171.2, as well as at δ (H) 2.19, indicated the presence of an AcO substitutent. The ¹H,¹H-COSY spectrum led to the identification of two partial structures: CH(1)–CH₂(2)–CH₂(3)–CH(4)–Me(15) and CH(4)–CH(5) (*Fig. 2*). Connections of these fragments to give a furanogermacrane skeleton were



Fig. 2. Key ${}^{1}H, {}^{1}H$ -COSY (—), HMBC (H \rightarrow C), and NOESY (H \leftrightarrow H) correlations of 2

revealed by HMBC correlations Me(14)/C(1), C(9), and C(10); H–C(9)/C(7) and C(8); Me(15)/C(3), C(4), and C(5); and H–C(5)/C(6), and C-atom signal at δ (C) 171.2 (*Fig.* 2). This established that the planar structure of **2** (*Fig.* 2) was very similar to that of **3**, except for the absence of MeO at C(2), and the replacement of C(1)=C(10) bond by 1,10-epoxy group [17]. The relative configuration of **2** was determined by NOESY correlation analysis. Specifically, correlations H_{β}–C(2)/Me(14) and Me(15), H–C(5)/Me(15); and H–C(1)/H_{α}–C(2) indicated that H–C(1), H–C(5), and Me(15) were α -, β -, and β -oriented, respectively. Accordingly, the structure of **2** was elucidated as 5 α -acetoxy-1,10-epoxyfuranogermacra-7,11-dien-6-one, and named myrrhasin A.

The eight known sesquiterpenoids from the resin of *C. opobalsamum* were identified as 5-acetoxy-2-methoxyfuranogermacr-1(10)-en-6-one (**3**) [17], (1(10)*E*,2*R*,4*R*)-2-methoxy-8,12-epoxygermacra-1(10),7,11-tetraen-6-one (**4**), [15][16], (1*E*)-3-methoxy-8,12-epoxygermacra-1,7,10,11-tetraen-6-one (**5**) [15], 7 α ,11-dihydroxycadin-10(14)-ene (**6**) [18], myrrhone (**7**) [19], 4 α ,10 α -dihydroxy-1 α ,5 α H-guaia-6-ene (**8**) [20], stigmasterol (**9**) [6], and cycloartan-24-ene-1 α ,2 α ,3 β -triol (**10**) [7] by comparison of their NMR data with those reported in the literature.

Compounds 1, 2, 6, and 8 were isolated by our group for the first time, and were evaluated for their cytotoxic activities against PC3 and DU145 human prostate tumor cell lines using the MTT (= 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2*H*-tetrazolium bromide) assay [21]. However, all of the tested compounds were inactive against both cancer cell lines ($IC_{50} > 50 \ \mu M$).

This work was supported by the *National Natural Science Foundation of China* (No. 81001376), *China Postdoctoral Science Foundation* (Nos. 201104602 and 20100471536), and *Postdoctoral Innovation Foundation of Shandong Province* (No. 201002018).

Experimental Part

General. Column chromatography (CC): silica gel (SiO₂, 200–300 mesh, 10–40 µm; Qingdao Haiyang Chemical Co. Ltd., China). TLC: Silica gel GF_{254} plates; visualization by heating the plates and spraying with 10% H₂SO₄/EtOH. Optical rotations: *GYROMAT-HP* polarimeter. 1D- and 2D-NMR spectra: *Bruker Avance 600* spectrometer at 600 (¹H) and 150 (¹³C) MHz; δ in ppm rel. to Me₄Si as internal standard, *J* in Hz. ESI-MS: *API 4000* mass spectrometer; in *m/z*. HR-ESI-MS: *LTQ-Orbitrap XL* mass spectrometer; in *m/z*.

Plant Material. The resin of *C. opobalsamum* was purchased in August 2010 from Jinan Jianlian Drug Store, P. R. China. It was imported from India and identified by Prof. *Lan Xiang*, School of Pharmaceutical Sciences, Shandong University, P. R. China. A voucher specimen (No. 20100817CO) has been deposited with the Department of Pharmacognosy, School of Pharmaceutical Sciences, Shandong University, P. R. China.

Extraction and Isolation. The powdered resins of C. opobalsamum (3.8 kg) were extracted by refluxing for 3 h with AcOEt (3×71) . After removal of AcOEt under reduced pressure, the extract (473 g) was suspended into H₂O, and partitioned with Petroleum (PE) and CH₂Cl₂. The CH₂Cl₂ extract (297 g) was subjected to CC (SiO₂; PE/AcOEt 100: $0 \rightarrow 0$:100) to give nine fractions, Fr. A – Fr. I. Fr. B (65 g) was separated by CC (SiO₂; PE/AcOEt 100:0 \rightarrow 90:10) to afford three subfractions, Fr. B1-Fr. B3. Fr. B1 was purified by CC (SiO₂; PE/AcOEt 50:1) to give Frs. B1a-B1c. Compound 3 (42 mg) was crystallized from Fr. Bla and purified by recrystallization (PE/Me₂O). Fr. Blb was subjected to HPLC (ODS; MeOH/H₂O 85:15) to give 5 (7 mg) and 7 (7 mg). Compounds 4 (5 mg) and 9 (6 mg) were purified from Fr. B1c by HPLC (ODS; MeOH/H2O 80:20). Fr. B3 was separated by CC (Sephadex LH-20; MeOH/CHCl₃ 1:1) to give Frs. B3a and B3b. Fr. B3a was submitted to CC (SiO₂; PE/AcOEt 40:1) and further purified by HPLC (ODS; MeOH/H₂O 80:20) to afford 2 (10 mg). Fr. C (16.1 g) was subjected to CC (SiO₂; PE/AcOEt 100:0 \rightarrow 85:15) to give two subfractions, Fr. C1 and Fr. C2. Compound 1 was obtained from Fr. C1 by HPLC (ODS; MeOH/H₂O 75:25. Fr. E (19.7 g) was separated by CC (SiO₂; (PE/AcOEt $100:0 \rightarrow 75:25$) to provide two subfractions, Frs. E1 and Fr. E2. Compound 6 (9 mg) and 8 (7 mg) were isolated from Fr. E2 by HPLC (ODS; MeOH/H₂O 73:27). Fr. F (27.2 g) was subjected to CC (SiO₂; PE/AcOEt $100: 0 \rightarrow 60: 40$) to give three subfractions, Fr. F1-Fr. F3. Compound 10 (14 mg) was purified from Fr. F2 by HPLC (ODS; MeOH/H₂O 68:32).

Myrrhanolide D (=(5\$,6\$,8\$,9R,11a\$)-8-*Methoxy*-3,6,10-*trimethyl*-2,4-*dioxo*-4,5,6,7,8,9-*hexahydro*-2H-9,11*a*-*epoxycyclodeca*[b]*furan*-5-*yl Acetate*; **1**). Yellow oil. $[\alpha]_{D}^{20}$ = +46.9 (c = 0.15, CHCl₃). ¹H- and ¹³C- NMR: see the *Table*. ESI-MS: 373.2 (20, $[M + Na]^+$), 368.3 (100, $[M + NH_4]^+$), 351.4 (60, $[M + H]^+$). HR-ESI-MS: 351.1454 ($[M + H]^+$, $C_{18}H_{23}O_7^+$; calc. 351.1444).

Myrrhasin A (=(4\$,5\$,10a\$)-4,7,10a-Trimethyl-6-oxo-1a,2,3,4,5,6,10,10a-octahydrooxireno[8,9]cyclodeca[1,2-b]furan-5-yl Acetate; **2**). Yellow oil. $[a]_{D}^{20} = -38.3$ (c = 0.26, CHCl₃). ¹H- and ¹³C- NMR: see the *Table*. ESI-MS: 329.5 (50, $[M + Na]^+$), 307.5 (100, $[M + H]^+$). HR-ESI-MS: 307.1552 ($[M + H]^+$, C₁₇H₂₃O⁺; calc. 307.1545).

REFERENCES

- [1] T. Shen, G.-H. Li, X.-N. Wang, H.-X. Lou, J. Ethnopharmacol. 2012, 142, 319.
- [2] E. El Ashry, N. Rashed, O. Salama, A. Saleh, *Pharmazie* 2003, 58, 163.
- [3] L. O. Hanuš, T. Řezanka, V. M. Dembitsky, A. Moussaieff, Biomed. Papers 2005, 149, 3.
- [4] T. Shen, H.-X. Lou, Chem. Biodiversity 2008, 5, 540.
- [5] R. Deng, Cardiovasc. Drug Rev. 2007, 25, 375.
- [6] T. Shen, L. Zhang, Y.-Y. Wang, P.-H. Fan, X.-N. Wang, Z.-M. Lin, H.-X. Lou, Bioorg. Med. Chem. Lett. 2012, 22, 4801.
- [7] T. Shen, W. Wan, H. Yuan, F. Kong, H. Guo, P. Fan, H. Lou, *Phytochemistry* 2007, 68, 1331.
- [8] T. Shen, H.-Q. Yuan, W.-Z. Wan, X.-L. Wang, X.-N. Wang, M. Ji, H.-X. Lou, J. Nat. Prod. 2008, 71, 81.
- [9] T. Shen, W.-Z. Wan, X.-N. Wang, H.-Q. Yuan, M. Ji, H.-X. Lou, *Helv. Chim. Acta* 2009, 92, 645.
 [10] T. Shen, W.-Z. Wan, X.-N. Wang, L.-M. Sun, H.-Q. Yuan, X.-L. Wang, M. Ji, H.-X. Lou, *Helv. Chim. Acta* 2008, 91, 881.
- [11] J.-L. Yang, Y.-P. Shi, Phytochemistry 2012, 76, 124.
- [12] J. Xu, Y. Guo, P. Zhao, C. Xie, D.-q. Jin, W. Hou, T. Zhang, Fitoterapia 2011, 82, 1198.
- [13] J. Xu, Y. Guo, P. Zhao, P. Guo, Y. Ma, C. Xie, D.-q. Jin, L. Gui, Fitoterapia 2012, 83, 801.
- [14] S.-L. Su, J. A. Duan, Y.-P. Tang, X. Zhang, L. Yu, F.-R. Jiang, W. Zhou, D. Luo, A.-W. Ding, *Planta Med.* 2009, 75, 351.
- [15] A. Dekebo, E. Dagne, O. Sterner, Fitoterapia 2002, 73, 48.

- [16] A. Dekebo, E. Dagne, L. K. Hansen, O. R. Gautun, A. J. Aasen, Tetrahedron Lett. 2000, 41, 9875.
- [17] C. H. Brieskorn, P. Noble, Tetrahedron Lett. 1980, 21, 1511.
- [18] H. I. El-Askary, M. R. Meselhy, A. M. Galal, *Molecules* 2003, 8, 670.
- [19] N. Zhu, S. Sheng, S. Sang, R. T. Rosen, C. T. Ho, Flavour. Frag. J. 2003, 18, 282.
- [20] C. Zhang, A. Zhou, M. Zhang, Zhongguo Zhongyao Zazhi 2009, 34, 994.
- [21] T. Mosmann, J. Immunol. Methods 1983, 65, 55.

Received August 29, 2013